

27 febbraio 2019  
Incontro del  
Gruppo di lavoro sui  
BIOFILM

Società dei Naturalisti in  
Napoli, via mezzocannone 8  
Ore 9.30

# **The invasion Model. A first step to model ecology in biofilms**

Maria Rosaria Mattei

Dipartimento di Matematica e Applicazioni *R. Caccioppoli*  
Università degli studi di Napoli *Federico II*

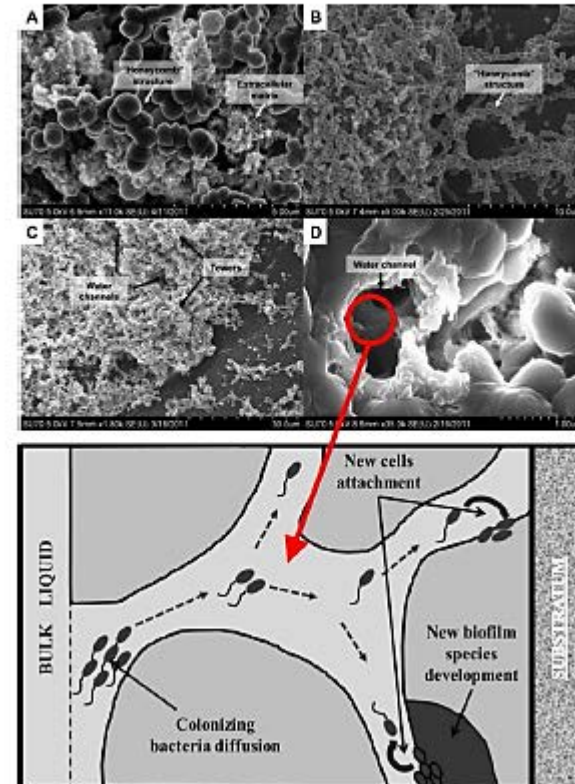
# Invasion in biofilms

infiltration of **planktonic cells** within the biofilm matrix and further establishment in **sessile form**

## Key features:

- *Ecological succession*
- Formation of *environmental microniches*
- Random movement or regulated cellular motility
- Rapid alterations in biofilm populations: *eradication or syntrophy*
- Biofilm reactors

$\psi_i$  planktonic bacteria  $\rightarrow X_i$  sessile bacteria

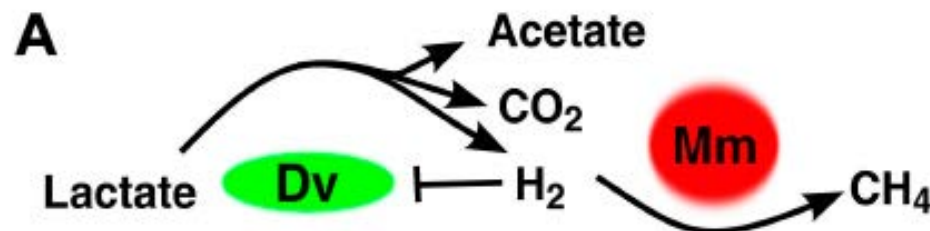


## MODEL APPLICATION 1

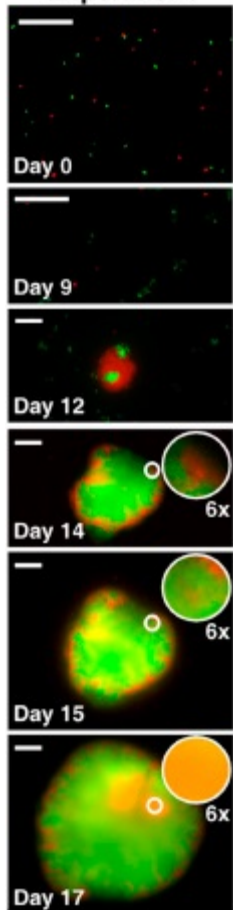
- Particulate components
  - $X_1$  Desulfovibrio vulgaris *DV*
  - $X_2$  Methanococcus maripaludis *MM*
- Dissolved components
  - $S_1$  Lactate
  - $S_2$  Acetate
  - $S_3$  Carbon Dioxide
  - $S_4$  Hydrogen
- Invading species
  - $\Psi_2$  invading bacteria  $\psi_{MM}$

**Obligatory cooperation through redox-coupling leads to partner intermixing.**

In the absence of sulfate and hydrogen, *DV* and *MM* cooperate through *redox coupling*. *DV* ferments Lactate and produces mainly Acetate,  $CO_2$ , and  $H_2$ .  $H_2$  is used by *MM* to reduce  $CO_2$  to methane.



## Experiment



RESEARCH ARTICLE



## Strong inter-population cooperation leads to partner intermixing in microbial communities

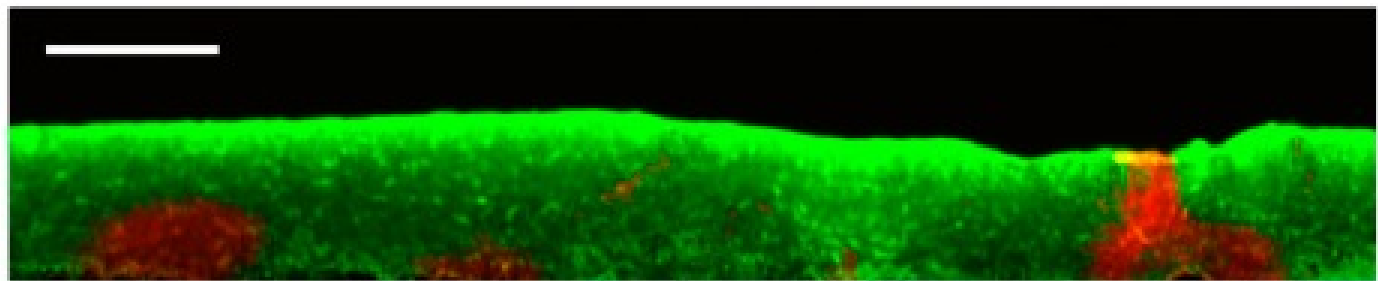
Babak Momeni<sup>1\*</sup>, Kristen A Brileya<sup>2</sup>, Matthew W Fields<sup>2</sup>, Wenying Shou<sup>1\*</sup>

<sup>1</sup>Division of Basic Sciences, Fred Hutchinson Cancer Research Center, Seattle, United States; <sup>2</sup>Department of Microbiology and Center for Biofilm Engineering, Montana State University, Bozeman, United States

**Abstract** Patterns of spatial positioning of individuals within microbial communities are often critical to community function. However, understanding patterning in natural communities is hampered by the multitude of cell–cell and cell–environment interactions as well as environmental variability. Here, through simulations and experiments on communities in defined environments, we

### EXPERIMENTAL OBSERVATIONS

- Biofilm Thickness  $L = 80\mu m$
- Biofilm Composition (Inner Layer) = 60% DV 40% MM
- Biofilm Composition (External Layer) = 80% DV 20% MM



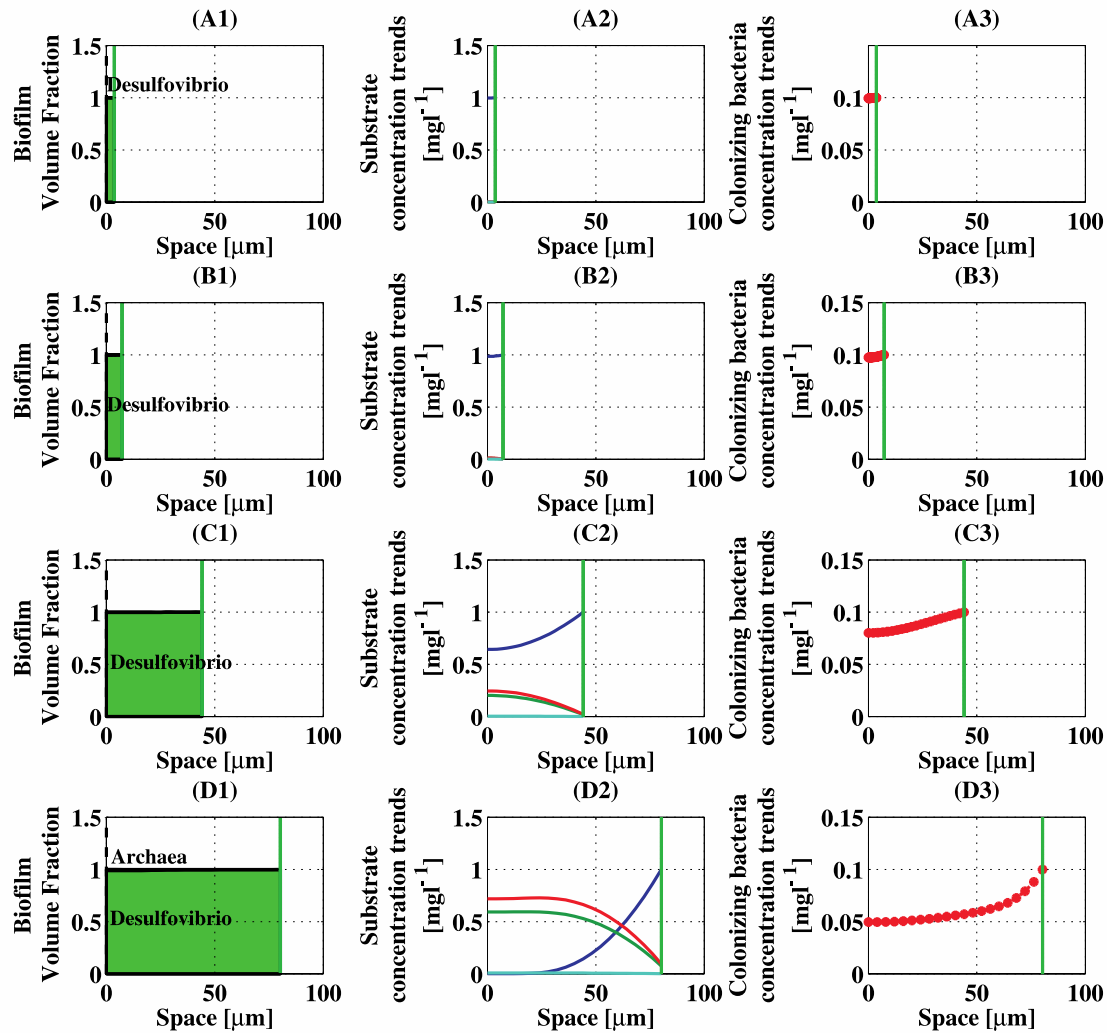


Figure: Effect of colonization on bacterial volume fractions, substrate concentration profiles, and  $\psi_2$  profile within biofilm after 1 (A1, A2, A3), 2 (B1, B2, B3), 5 (C1, C2, C3), 10 (D1, D2, D3) days. Blue line - Lactate; red line - Acetate; green line - Carbon dioxide; ciano line - Hydrogen.

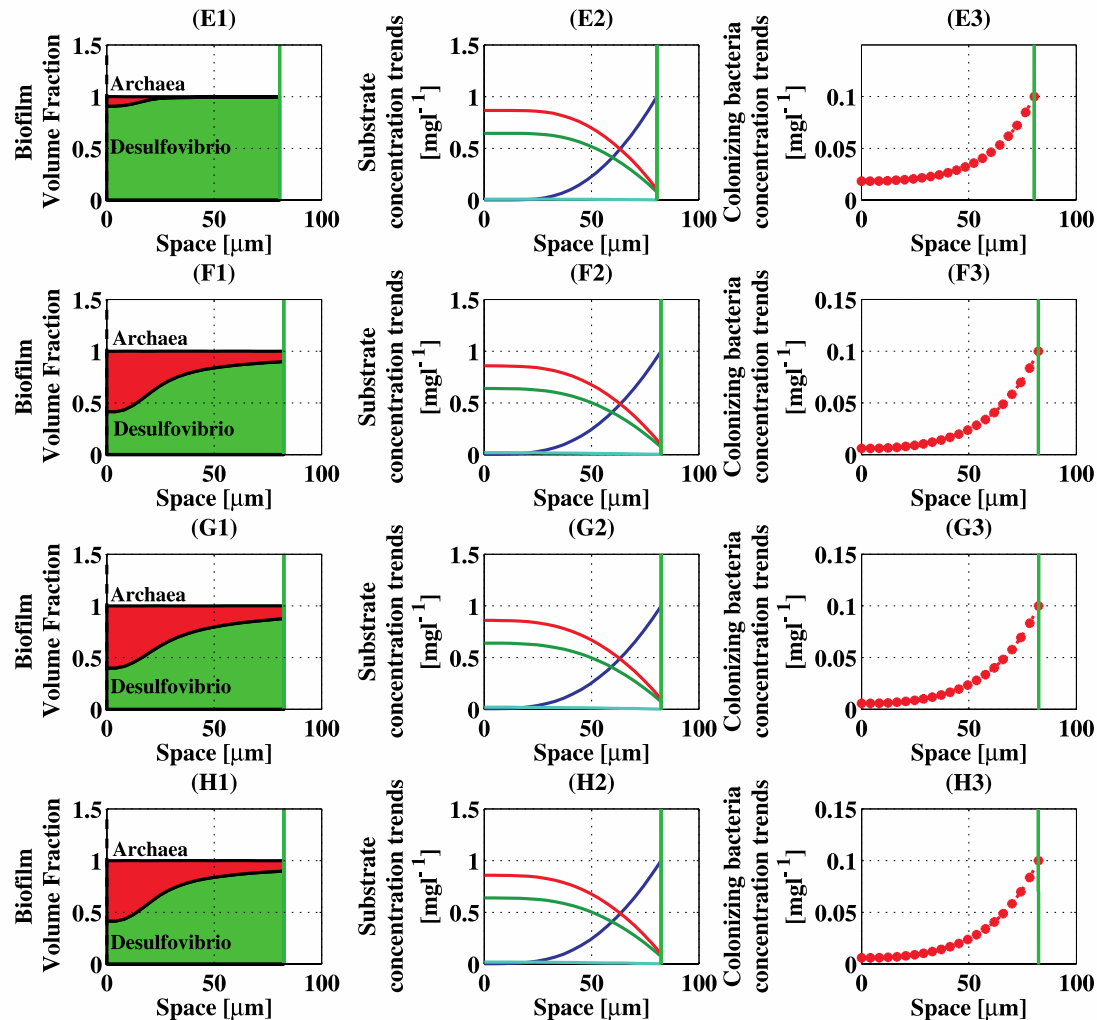


Figure: Effect of colonization on bacterial volume fractions, substrate concentration profiles, and  $\psi_2$  profile within biofilm after 20 (E1, E2, E3), 30 (F1, F2, F3), 50 (G1, G2, G3), 90 (H1, H2, H3) simulation days. Blue line - Lactate; red line - Acetate; green line - Carbon dioxide; ciano line - Hydrogen

## MODEL APPLICATION 2

## ● Particulate components

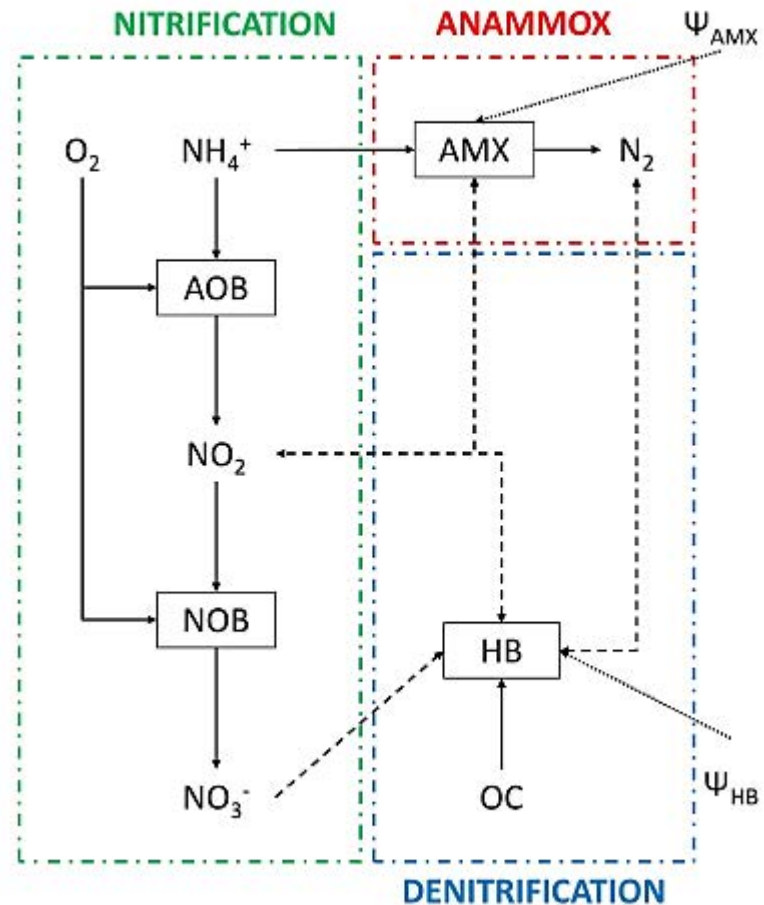
- $X_1$  Ammonium Oxidizing Bacteria *AOB*
- $X_2$  Anaerobic Ammonium Oxidizing Bacteria *AMX*
- $X_3$  Nitrite Oxidizing Bacteria *NOB*
- $X_4$  Heterotrophic Bacteria *HB*
- $X_5$  Inert material *I*

## ● Dissolved components

- $S_1$  Ammonium  $S_{NH4}$
- $S_2$  Nitrite  $S_{NO2}$
- $S_3$  Nitrate  $S_{NO3}$
- $S_4$  Organic Carbon  $S_{OC}$
- $S_5$  Oxygen  $S_{O2}$

## ● Invading species

- $\Psi_2$  invading bacteria  $\psi_{AMX}$
- $\Psi_4$  invading bacteria  $\psi_{HB}$





## Simulation 2 - AMX &amp; HB invasion

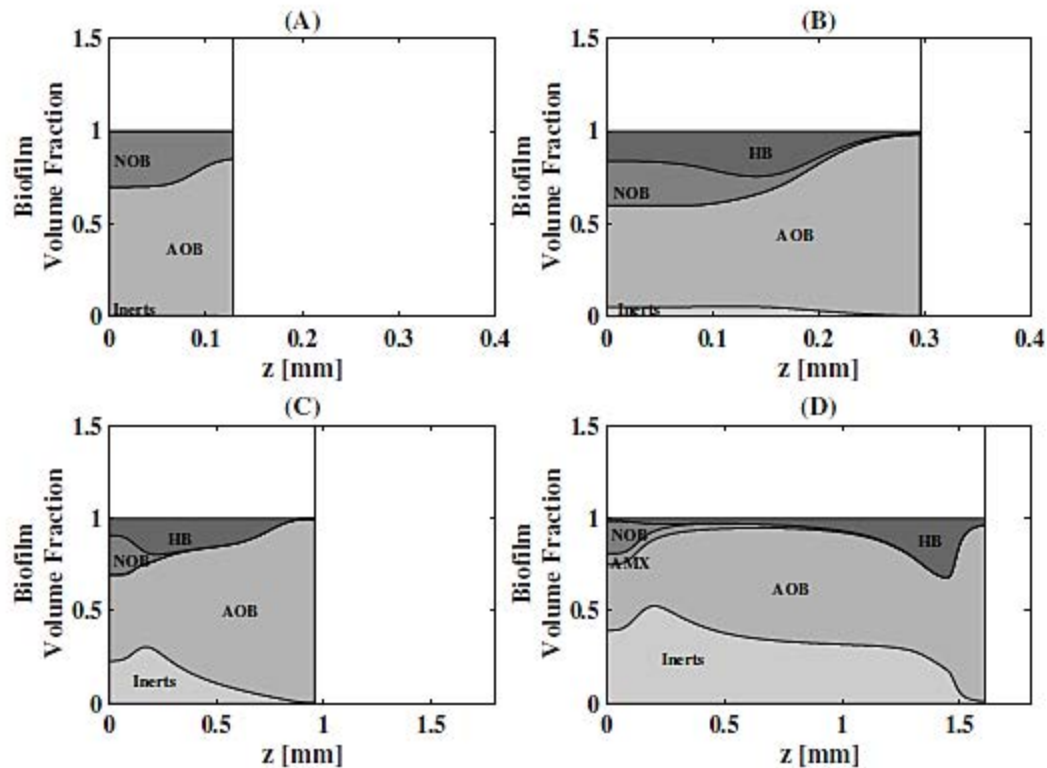


Figure: Microbial species distribution of a multispecies biofilm undergoing  $\psi_2$  and  $\psi_4$  colonization after 2(A), 5(B), 20(C), 50(D) days simulation time. The substratum is placed at  $z = 0$ .